Antimicrobial effect of dried sage on the microbiological state of fresh Hungarian sausage

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SUMMARY

The purpose of this study was to evaluate the microbial effect of dried sage (Salvia officinalis L.) on the traditional Hungarian sausage. We added 0.5, 1, 1.5, and 2 w/w% of sage to the sausages and tested them on the 0^{th} , 7^{th} and 14^{th} day. The added dried sage had no effect on the tested microorganisms, but the sage extract inhibited Salmonella, Enterococcus faecium and Staphylococcus aureus.

Keywords: sausage; sage; antimicrobial effect

INTRODUCTION

Throughout human history, sage has been a wellknown herb; it was used by the Chinese to develop drugs, or by the Egyptians, Greeks and Romans as a herb and drug. Salvia officinalis was one of the species which has pharmacological effects. The plant has biological activities, for example virustatic, fungistatic, and antibacterial effects. The biological effects of sage depend on the content of camphor, 1,8-cineole, α thujone, and β -thujone. (Hamidpour et al., 2014) Sage has antibacterial activity through 1.8-cineole, camphor, thujone against Bacillus subtilis, Bacillus cereus. Two other sage components, oleanolic acid and ursolic acid, can also inhibit different pathogenic bacteria such as penicillin-resistant Streptococcus pneumonia, and methicillin-resistant Staphylococcus aureus. Shirazi et al. (2008) tested in vitro the antibacterial effect of sage extract against Salmonella typhi, Salmonella flexneri, vulgaris. Salmonella sonnei. Pseudomonas Staphylococcus aureus, enterotoxigenic Escherichia coli (ETEC), and Pseudomonas aeruginosa. They found that sage extract had no effect on P. aeroginosa, and ETEC, but it inhibited P. vulgaris, S. flexneri, and S. sonnei. Moreover, it had as strong effect as ampicillin and streptomycin against S. typhi. The effect against S. aureus was as strong as a wide scale of antibiotics used against S. aureus such as vancomycin (Shirazi et al., 2008; Ghorbani and Esmaeilizadeh, 2017). The active components of plants are usually hydrophobic by nature. Because of this attribution their primary target is the cell membrane. They can cause loss of ions, enzymes and metabolites from the cell which will lead to the death of the cell (Mitić-Ćulafić et al., 2005).

Císarová et al. (2018) tested α -thujone and β thujone, 1,8-cineole, borneol, sage extract, and sage essential oil with *E. coli, K. pneumoniae, E. faecalis,* and *S. aureus*. They found α -thujone and β -thujone could not inhibit these strains. *E. faecalis* was resistant to the sage essential oil, while 1-8-cineol had no effect on *E. coli* and *S. aureus*. The sage extract had a strong antibacterial effect in case of *E. faecalis* and *K. pneumoniae* (Císarová et al., 2018). There are sprays with added herbal active ingredient used by the food industry. They have several advantages, like easy handling, short contact with the medium, good reproducibility. These sprays can only be used on the surface of the product. Maybe an extract used in the product could have more advantages, for example it has a positive effect on the taste of the product, and the appearance of the product. The food industry is on the development of healthier products in the past years, one of the additive group is the synthetic antioxidants. They delay or inhibit for example oxidation in foods. The main point is the usage of plants and their active components, which are of natural origin (Pavlić et al., 2017; Munekata et al., 2020).

One of the most common meat products is the fresh pork sausage, which has several types depending on the country or region of origin, the applied spices and herbs. Fresh pork sausages have high water activity, they are not heat treated until the consumption so the basic microbial flora of fresh meat has effect on the shelf life of the product. The food industry is trying to replace the artificial preservatives with agents of natural origin. Natural agents have disadvantages, they have higher price than artificial preservatives and they are not as effective as the artificial ones (Šojić et al., 2018).

MATERIALS AND METHODS

The minced pork meat was obtained from a local meat product factory, Darnó-hús (Funkcio Ltd, Darnózseli, Hungary), the used spices were obtained from commercial market (Kotányi Hungary Spices Processing and Sales Ltd., Törökbálint, Hungary; Lacikonyha Hungary Ltd., Budapest, Hungary). The used dried and ground sage was harvested from the parcel of the Széchenyi István University, Faculty of Agricultural and Food Sciences, Department of Food Science.

Preparation of the pork sausage

The raw pork meat was purchased in a ground form. We added the needed spices according to the recipe of the traditional Hungarian sausage: sweet paprika



DOI: 10.34101/ACTAAGRAR/1/8708

powder (1.88 w/w%), salt (1.88 w/w%), black pepper (0.38 w/w%), white pepper (0.19 w/w%), cumin powder (0.38 w/w%), nutmeg powder (0.09 w/w%), and freshly ground garlic (1.40 w/w%), then we mixed it and 1–1 kilograms were measured into plastic boxes. We added 0.5, 1.0, 1.5 and 2.0 w/w% of sage to the pulp and mixed them. One of the batter did not contain sage powder, that was the control. The batters were stuffed into natural casing (pig small intestines, Böllérbolt, Pécs, Hungary) and then were vacuum packed into 150x200 vacuum bags (Gasztronauta Ltd., Győr, Hungary). The core temperature of the meat and the pulp did not reach 5 °C. We kept the sausages at 4 °C until we performed the tests. The chilled products were tested without heat treatment.

Extraction of the sage

We extracted the sage according to Durling et al. (2007). We ground (Hausmeister Ltd., Norfolk, United Kingdom) the sage that was previously dried in a drying oven for 24 h on 35 °C. We prepared the extraction solution by using 99.8% ethanol and deionised water. The solution contained 81% ethanol, the solvent-to-sage ratio was 6:1. We kept the extraction at 40 °C for 4 h. To remove the ethanol from the solution we used drying oven for 3 h at 70 °C, the leftover solution was poured into vials and kept under 4 °C until the tests were evaluated.

Microbiology analysis

The sausages were tested on the 0th, 7th and 14th days. The samples were tested for *Staphylococcus aureus* on Baird Parker (BP, Biolab Ltd, Budapest, Hungary) agar at 37 °C, 48 h, for yeast and mold on Yeast Glucose Chloramphenicol (YGC, Biolab Ltd, Budapest, Hungary) agar at 25 °C, 120 h, for *Clostridium perfringens* on Tryptose Sulfite Cycloserine (TSC, Biolab Ltd, Budapest, Hungary) agar at 37 °C, 48 h, *coliforms* and *Eschericia coli* on ChromoCult Coliform (CC, Biolab Ltd, Budapest, Hungary) agar at 37 °C, 48 h, and total plate count on Plate Count (PC, Biolab Ltd, Budapest, Hungary) agar

at 30 °C, 72 h. In case of *Salmonella* the sample was first enriched in Buffered Pepton Water (Biolab Ltd, Budapest, Hungary), then selectively enriched in Rappaport-Vasiliadis broth (RV, Biolab Ltd, Budapest, Hungary), and finally one more selective enrichment was done on Xylose Lysine Desoxychoate agar (XLD, Biolab Ltd, Budapest, Hungary).

Agar well diffusion test

The extracted sage solution was tested for inhibition for *Salmonella, E. coli, S. aureus, and E. faecium* on Tryptic Soy Agar (Biolab Ltd, Budapest, Hungary) at 37 °C for 24 h. After the incubation time we measured the inhibition zones around the drilled holes.

RESULTS AND DISCUSSION

We tested the sausages on the 0th, 7th and 14th days. Vacuum packed sausages do not have a longer shelf life than 14 days because of the microbiological and enzyme activity.

On the first and last test round, we did not find typical *C. perfringens* colonies on the surface of the agar. On the 7th day in case of 0% of sage $1.1*10^3$, in case of 0.5% $1.1*10^3$ and in case of 1% of sage $6.8*10^2$ CFU/g were detected in the experimental groups, respectively. According to the Decree No. 4 of 1998 of the Ministry of Health the limit for *C. perfringens* is 10^2 CFU/g, consequently, on the 7th day sausages it reached and exceeded the limit.

In the case of *S. aureus* we found black colonies on the BP agar, but they did not have the typical clear zone around the colonies which characterize the typical *S. aureus* colonies on BP agar caused by the egg yolk tellurite emulsion (Biolab Ltd, Budapest, Hungary). The confirmatory test was evaluated by Staphytect Plus (Thermo Fisher Scientific Inc, Waltham, USA), but we did not detect agglutination.

In the test of *E. coli* the colonies showed an increase until the 7th day, and then a decrease was observed on the 14th day (*Table 1*). The Decree No. 4 of 1998 of the Ministry of Health does not regulate the limit of *E. coli* in fresh sausages.

days	Percentage of sage						
	0	0.5	1	1.5	2		
0	2.9*10 ¹	$3.8*10^{1}$	$5.8*10^{1}$	7,1*10 ¹	5.4*10 ¹		
7	5*10 ¹	$1.3*10^{2}$	$4.4*10^{2}$	$8.8*10^{1}$	$6.2*10^{2}$		
14	$1.7*10^{1}$	$8.3*10^{1}$	$1.3*10^{2}$	$0.8*10^{1}$	$1.2*10^{2}$		

Table 1. Microbial results of the Escherichia coli test (CFU/g)

In the case of coliforms nearly ten times more CFU/g were found compared to the *E. coli* tests (*Table 2*). The number of coliforms increased until the 7th day as well, whereas on the last day the number nearly

approached the initial values of day 0. The Decree No. 4 of 1998 of the Ministry of Health does not regulate the limit of coliforms in meat products.



DOI: 10.34101/ACTAAGRAR/1/8708

days	Percentage of sage (%)						
	0	0.5	1	1.5	2		
0	$5.1*10^{2}$	$4.8*10^{2}$	$4.5*10^{2}$	$4.1*10^{2}$	$3.6*10^{2}$		
7	$3.7*10^{3}$	$2.5*10^{3}$	$8.4*10^{3}$	$1.2*10^{4}$	$8.6*10^{3}$		
14	$4.3*10^{2}$	$5.7*10^{2}$	$6.2*10^{2}$	$6.5*10^{2}$	$5.9*10^{2}$		

Table 2. Microbial results of coliforms (CFU/g)

In the case of yeast the sausage which did not contain sage had less than 10 CFU/g of yeast colonies (*Table 3*). Colonies showed rising only on the 7th day. Sausages that contained sage had a 10^2 initial yeast number but they reached less than 10 on the 14th day. Only the 2% sage product had increasing numbers until the 7th day, but it reached less than 10 on the 14th day,

as well. We could not find yeast colonies on the agar plates.

In the case of mold their number showed a halving in case of the 7th day test (*Table 4*). On the 14th day we could not find typical colonies on the agar plates. We also tested the sage, it had $4.5*10^2$ initial yeast number.

Table 3. Microbial results of the yeast test (CFU/g)

days	Percentage of sage (%)					
	0	0.5	1	1.5	2	
0	$< 10^{1}$	$4.05*10^{2}$	$4.1*10^{2}$	$3.3*10^{2}$	5.7*10 ²	
7	$3.1*10^{2}$	$2.6*10^{2}$	$1.5*10^{2}$	$2.5*10^{2}$	$7*10^{2}$	
14	$< 10^{1}$	<101	$< 10^{1}$	$< 10^{1}$	<101	

Table 4. Microbial results of mold test (CFU/g)

days	Percentage of sage (%)						
	0	0.5	1	1.5	2		
0	$5.6*10^{2}$	$2.8*10^{2}$	$2.3*10^{2}$	$2.4*10^{2}$	$4.5*10^{2}$		
7	$2.4*10^{2}$	$1.5*10^{2}$	$1.3*10^{2}$	$1.6*10^{2}$	$1.5*10^{2}$		
14	$< 10^{1}$	$< 10^{1}$	$< 10^{1}$	<101	$< 10^{1}$		

In the case of the total plate count we did not find the same results like in the case of other tested microorganisms (*Table 5*). We found only increasing, in three cases, the sausages which contained 1%, 0.5%, and 1.5% on the 14th day had decreasing. The logical result would be a decrease in the case of 2% of sage because it contained the most sage, it had the highest active component content but we found the highest increase in the case of 2%. We tested the raw meat too, it contained $1.7*10^5$ microorganisms on the 0^{th} day. The increase was three and in one case four order of magnitude. Without the vacuum packaging these number could have been reached within few days and not in two weeks.

	days			Percentage of sage (%)		
	0	0.5	1	1.5	2	
0	$2.9*10^{3}$	$2.4*10^{3}$	$2.4*10^{3}$	$1.9*10^{3}$	$1.4*10^{3}$	
7	$3.6*10^{6}$	$3.01*10^{6}$	$5*10^{6}$	$4.6*10^{6}$	$3.9*10^{6}$	
14	$7.3*10^{6}$	$2.2*10^{6}$	9.3*10 ⁵	$2.7*10^{6}$	$1.54*10^{7}$	

Agar well diffusion test

Our sage extract did not have inhibitory effect on *E. coli*, the solution diffunded into the agar but we did not observe clear inhibition. The solution had inhibition for *E. faecium*, *S. aureus* and *Salmonella*. In case of *E. faecium* the average diameter was 7.7 mm. The effect of the extract on *S. aureus* resulted in the clearest inhibition, in the average diameter of 10 mm. The strongest inhibition was in the case of *Salmonella*, the average diameter was 20.3 mm.



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CONCLUSIONS

The application of herb extracts is a new but rapidly developing trend in the food industry. The fresh or dry parts of herbs do not contain as much active components such as extracts like essential oils. The difference of the effects is clearly visible from the articles which tested the effects of essential oils against dried herbs. Dried herbs are needed because consumers are buying food products "with their eyes", too, therefore a product containing only essential oil will not have the same effect on the consumer like a product with essential oil and herb pieces.

The next studies will be about the essential oils and their effects on the microbial state and

organoleptic properties of Hungarian fresh and riped sausages.

ACKNOWLEDGEMENTS

Supported by the ÚNKP-20-3-I, New National Excellence Program of the Ministry for Innovation and Technology from the source of the National Research, Development and Innovation Fund.

This work was supported by grant number EFOP-3.6.3-VEKOP-16-2017-00008 entitled "Innovative Scientific Institutions in Domestic Agricultural Higher Education". The project is co-financed by the European Union and the European Social Fund.

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